Discrete-Time Model Representation for Biochemical Pathway Systems

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Abstract—This paper presents the importance of formulating general discrete-time model representations for current pathway deterministic modeling study. Discrete-time models can be considered as a link between continuous-time kinetic reactions and discrete-time experimentation as well as computer based simulation and analysis. In the paper, different discretization techniques are investigated according to different sorts of ODE model structures. Two discretization strategies are mainly focused, that are one-step Taylor/Lie series based method and multi-step Runge-Kutta method. A new discretization is proposed for bilinear in states pathway models. Finally, the superiority of using Runge-Kutta based approach as general discrete-time model representations are concluded.

Index Terms—Biochemical pathway modeling, discretization, sensitivity analysis, systems biology.

I. INTRODUCTION

Based on large amounts of experiments, traditional biochemists and molecular biologists have developed many qualitative models and hypothesis for biochemical pathway study [1]-[3]. However, in order to evaluate the completeness and usefulness of a hypothesis, produce predictions for further testing, and better understand the interaction and dynamic of pathway components, qualitative models are no longer adequate. There has recently been a focus on more quantitative approach in systems biology study. In the past decade, numerous approaches for quantitative modeling of biochemical dynamics are proposed, such as Robert [4], Anand and Douglas [5], Jeff et al. [6], Robert and Tom [7], Tyson et al. [8], Wolkenhauer [9] [10], etc. Among these approaches, the most prominent method is to use ordinary differential equations (ODEs) to model biochemical reactions according to mass action laws. It should be noted that using ODEs to model biochemical reactions assumes that the system is well-stirred in

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a homogeneous medium and that spatial effects, such as diffusion, are irrelevant, otherwise partial differential equations (PDEs) should be used [11]. In the literature, almost all publications related to pathway modeling are based on continuous-time ODE model representations. Using continuous-time ODEs facilitate researchers on analytical study and understanding, whereas it also brings difficulties for numerical computation and computer based simulation. Therefore, constructing the corresponding discrete-time model representations is particular important in systems biology study.

There are many reasons to formulate discrete-time model representations in pathway modeling research. Firstly the real biochemical kinetic reactions take place in continuous time, whereas experimental data are measured by sampling the continuous biochemical reaction outputs, and computer based analysis and simulation all depend on discrete-time datasets. Therefore, a discrete-time model could be an interface between real kinetic reaction, experimentation and computer based simulation. A delicate discrete-time model can not only assist people to better understand pathway reaction dynamics and reproduce the experimental data, but also generate predictions for computer based analysis which leaves out the expensive and time consuming experiment process. Moreover, it can be a crucial tool for further experimental design study, such as state measurement selection and sampling time design. Secondly, as we know parameter estimation is an active and important research topic in systems biology study. Estimating parameters of continuous ODEs is usually a computational costly procedure, as even for linear ODEs it is a non-quadratic problem. Whereas optimization when considering discrete-time based models, although we can not change the fundamental nature of the optimization process, however, an iterative polynomial discrete-time model could possibly simplify the structure of continuous ODEs, especially for some nonlinear cases. This can help researchers to develop new parameter estimation approaches based on the discrete-time models. Furthermore, dynamic sensitivity analysis plays an important role in parameter selection and uncertainty analysis for system identification procedure [12], in practice, local sensitivity coefficients are obtained by solving the continuous sensitivity functions and model ODEs simultaneously. As sensitivity functions are also a set of ODEs with respect to sensitivity coefficients, it would be worthwhile to calculate sensitivity coefficients in a similar discrete-time iterative way.

In practice, there are several methods can be considered for discretization of ODEs. One sort of methods is based on Taylor

or Lie series expansion which is one-step-ahead discretization stratagem. For models represented by linear ODEs which means linear in states, the discrete-time model representation is given in discrete-time control system textbooks [13] as linear ODEs can be expressed as state-space equations. Unfortunately, for nonlinear ODEs there is no such general direct discretization mapping. In mathematical and control theory, some discretization techniques related to nonlinear ODEs are comparatively reviewed and discussed in section 3. Considering real biological signaling cases, we investigate a time varying linear approach for bilinear in states ODEs situation based on Taylor expansion and Carleman linearization. However, even for this method the mathematical model expression would be complex when considering higher order approximation. Another important discretization strategy discussed in this paper is multi-step discretization approach based on Runge-Kutta method. One advantage of this approach is it improves the discretization accuracy by utilizing multi-step information for approximation of one-step-ahead model prediction. Moreover, it gives a general exact discrete-time representation for both linear and nonlinear biochemical ODE models.

II. CONTINUOUS-TIME PATHWAY MODELING

A. Continuous-Time Model Representation

In the literature [14] [15], signal pathway dynamics can usually be modeled by the following ODE representation:

$$\begin{aligned} \dot{\mathbf{x}}(t) &= \mathbf{f}(\mathbf{x}(t), \mathbf{u}(t), \mathbf{\theta}), \quad \mathbf{x}(t_0) = x_0 \\ \mathbf{y}(t) &= \mathbf{g}(\mathbf{x}(t)) \end{aligned} \tag{1}$$

where $\mathbf{x} \in \mathbb{R}^{m}$, $\mathbf{u} \in \mathbb{R}^{p}$, and $\boldsymbol{\theta} \in \mathbb{R}^{n}$ are the state, input and parameter column vectors. x_{0} is the initial states vector at t_{0} . From biochemical modeling viewpoint, \mathbf{x} represents molecular concentrations; \mathbf{u} generally represents external cellular signals; $\boldsymbol{\theta}$ stands for reaction rates. $\mathbf{f}(\cdot)$ is a set of linear or nonlinear functions which correspond to a series of biochemical reactions describing pathway dynamics. $\mathbf{g}(\cdot)$ here is the measurement function which determines which states can be measured. For the simplest case, if all the states can be measured, the measurement function \mathbf{g} in (1) is an identity matrix. Otherwise, \mathbf{g} is a rectangular zero-one matrix with corresponding rows with respect to unmeasurable states deleted from the full rank identity matrix \mathbf{I}_{m} .

When model ODEs are linear and time invariant in states, which is also known as linear ODEs, (1) can be simplified as:

$$\mathbf{x}(t) = \mathbf{A}\mathbf{x}(t) + \mathbf{B}\mathbf{u}(t)$$

$$\mathbf{y}(t) = \mathbf{g}(\mathbf{x}(t))$$
 (2)

Here $\mathbf{A}_{m \times m}$ is the parameter matrix, $\mathbf{B}_{m \times p}$ is the known input matrix. For most systems biology pathway models **A** is typical a sparse matrix, only the corresponding reaction state terms appear in the forcing function. However, this kind of linear ODEs is not prevalent for representation of most biochemical reactions, only if it is an irreversible chain reaction. An illustrative example is the first-order isothermal liquid-phase chain reaction [16] [17]:

$A \xrightarrow{\theta_1} B \xrightarrow{\theta_2} C$

It states from liquid reaction component A to liquid product B and then to liquid product C. This reaction process can be model by following ODEs:

$$\begin{aligned} \dot{x}_1 &= -\theta_1 x_1 \\ \dot{x}_2 &= \theta_1 x_1 - \theta_2 x_2 \end{aligned} \tag{3}$$

where x_1, x_2 denote concentrations of components *A* and *B*, which were the only two concentrations measured. Therefore, component *C* does appear in the model. This ODE model (3) can be readily represented as linear state space form (2).

More generally, and more applicably, we can consider ODEs that are linear in their unknown parameters, but not necessarily states. For instance, the Michaelis-Menten enzyme kinetics [20] [21], JAK-STAT [18], ERK [19], $TNF\alpha - NF - \kappa B$ [20] and $I\kappa B - NF - \kappa B$ [15] pathway models are all bilinear in the states but linear in parameters. The state function of this kind of ODEs can be represented as:

$$\dot{\mathbf{x}}(t) = \mathbf{F}(\mathbf{x}(t))\mathbf{\Theta} \tag{4}$$

where $\mathbf{F}(\cdot)$ represents a set of nonlinear functions which is also commonly presence as a sparse matrix. For example, considering a bilinear in states model, when a reaction only takes place in the presence of two molecules, most elements of the corresponding row in \mathbf{F} would be zeros except for those related to this two states. Here we do not take account of model inputs $\mathbf{u}(t)$ as most of these published pathways are not considered subject to external cellular signals.

B. Parameter Estimation

Given the model structure and experiment data of measured state variables, the aim of parameter estimation is to calculate the parameter values that minimize some loss function between the model's prediction and measurement data. Considering the set of $\{\tilde{y}_i(k)\}_{i,k}$ as the measurement data and the corresponding model's predictions $\{y_i(k, 0)\}_{i,k}$, which is simply discrete-time sampling of continuous-time ODE model's output $\mathbf{y}(t)$, then a standard least squares loss function along the trajectory gives:

$$\hat{\boldsymbol{\theta}} = \arg\min\frac{1}{2}\sum_{i}\sum_{k}\omega_{i}(\tilde{y}_{i}(k) - y_{i}(k,\boldsymbol{\theta}))^{2}$$
(5)

where the double sum can be taken simply as taking the expected value over all states (*i*) and over the complete trajectory (*k*), ω_i are the weights to normalize the contributions of different state signals and can be taken as

$$\omega_i = \left(1/\max_{\nu}(\tilde{y}_i(k))\right)^2 \tag{6}$$

We assume that the model hypothesis space includes the optimal model, so that when $\hat{\boldsymbol{\theta}} = \boldsymbol{\theta}^*$, the model's parameter are correct and $y_i(k, \hat{\boldsymbol{\theta}}) = y_i^*(k)$. Typically, we assume that the states are not directly measurable, but are subject to additive measurement noise:

$$\tilde{y}_i(k) = y_i^*(k) + \aleph(0, \sigma_i^2) \tag{7}$$

Here, the noise is zero mean Gaussian, where the variances depend on the state.

We have to take into account the fact that we can only measure/estimate the states rather than the first-order derivative of states to time (left-hand-side of ODEs), so even for linear ODE models, the optimization problem is not quadratic. In the literature, parameter estimation of pathway ODEs is usually reduced into solving nonlinear boundary value problem using multiple shooting method [22]-[24].

C. Sensitivity Analysis

Dynamic sensitivity analysis plays an important role in parameter selection and uncertainty analysis for system identification procedure. The first-order local sensitivity coefficient $s_{i,j}$ is defined as the partial derivative of *i*th state to *j*th parameter

$$s_{i,j}(t) = \frac{\partial x_i(t)}{\partial \theta_i} = \frac{x_i(\theta_j + \Delta \theta_j, t) - x_i(\theta_j, t)}{\Delta \theta_i}$$
(8)

In (8), sensitivity coefficient is calculated using finite difference method (FDM), however, the numerical values obtained may vary with $\Delta \theta_j$, and repeated measurement of state is required at least once for each parameter. In practice [15], direct differential method (DDM) is employed as an alternative by taking partial derivative of (1) with respect to parameter θ_j , and the absolute parameter sensitivity equations can be written as:

$$\frac{d}{dt}\frac{\partial \mathbf{x}}{\partial \theta_j} = \frac{\partial \mathbf{f}}{\partial x}\frac{\partial \mathbf{x}}{\partial \theta_j} + \frac{\partial \mathbf{f}}{\partial \theta_j} \Leftrightarrow \dot{S}_j = J \cdot S_j + P_j, \quad S_j(t_0) = S_0 \quad (9)$$

where J and P_j are the Jacobian matrix and parameter Jacobian matrix. By solving the *m* equations in (1) and $n \times m$ equations in (9) together as a set of differentical equations, both $\mathbf{x}(t)$ and $\mathbf{S}(t)_{m \times n}$ can be determined simultaneously.

For ODEs are linear in both parameters and states, a special case described in (2), and when assuming biochemical reactions are autonomous which means do not affected by external inputs \mathbf{u} , the corresponding linearized sensitivity equations can be expressed as:

$$\dot{\mathbf{S}}(t) = \mathbf{A}\mathbf{S}(t) + \mathbf{P}(t) \tag{10}$$

where **P** is the $m \times n$ parameter Jacobian matrix.

For linear in the parameters ODEs (4), the corresponding sensitivity equations can be simplified as:

$$\dot{\mathbf{S}}(t) = \frac{\partial (\mathbf{F}(\mathbf{x}(t)\mathbf{\theta})}{\partial \mathbf{x}} \mathbf{S}(t) + \mathbf{F}(\mathbf{x}(t))$$
(11)

Parameter sensitivity coefficients provide crucial information for parameter importance measurement and further parameter selection. A measure of the estimated parameters' quality is given by the Fisher information matrix (FIM):

$$\mathbf{F} = \sigma^2 \left(\frac{d\mathbf{x}}{d\mathbf{\theta}}\right)^T \left(\frac{d\mathbf{x}}{d\mathbf{\theta}}\right) = \sigma^2 \mathbf{S}^T \mathbf{S}$$
(12)

which is a lower bound on the parameter variance/covariance matrix. This is a key measure of identifiability which determines how easily the parameter values can be reliably estimated from the data, or alternatively, how many experiments would need to be performed in order to estimate the parameters to a pre-defined level of confidence.

In the literature, several algorithms for parameter selection are proposed based on parameter sensitivity analysis [26] [27]. Besides, many optimal experimental design methods [27]-[29] are developed based on maximize the information of FIM according to commonly used optimal design criteria [30].

III. DISCRETE-TIME MODEL REPRESENTATION

Equation (1) describes pathway dynamics in continues time. However, in real experiment the measurement results are obtained by sampling continues time series, and later on system analysis, parameter estimation and experimental design are all based on these discrete data sets. Therefore, it's important to formulate a discrete time model representation.

A. One-Step System Discretization

For linear in the states ODEs, the exact discrete-time representation of system ODEs (2) will take the form:

$$\mathbf{x}(k+1) = \mathbf{G} \cdot \mathbf{x}(k) + \mathbf{H} \cdot \mathbf{u}(k)$$
(13)

If we denote t = kT and $\lambda = T - t$, where,

$$\mathbf{G} = e^{\mathbf{A}T}, \quad \mathbf{H} = \left(\int_0^T e^{\mathbf{A}\lambda} d\lambda\right) \mathbf{B}$$
(14)

If matrix A is nonsingular, then H given in (14) can be simplified to

$$\mathbf{H} = \left(\int_0^T e^{\mathbf{A}\lambda} d\lambda\right) \mathbf{B} = \mathbf{A}^{-1} (e^{\mathbf{A}T} - \mathbf{I}) \mathbf{B} = (e^{\mathbf{A}T} - \mathbf{I}) \mathbf{A}^{-1} \mathbf{B}$$
(15)

Similarly, if the biochemical reactions are autonomous, the discrete-time sensitivity equations can be written as:

$$\mathbf{S}(k+1) = \mathbf{G} \cdot \mathbf{S}(k) + \mathbf{B}_d \mathbf{P}(k)$$
(16)

where,

$$\mathbf{B}_{d} = \int_{0}^{T} e^{\mathbf{A}\lambda} d\lambda \tag{17}$$

The discrete-time representation discussed above had been mentioned in some pathway modeling literature [31] [32].

Unfortunately, there is no general exact mapping between continuous and discrete time systems when ODEs are nonlinear. In numerical analysis and control theory, there are several methods had been discussed for one-step discretization of nonlinear ODEs. Such as finite differences method [33], which comprises Euler's method and finite order Taylor series approximation, Carleman linearization [34], Jacobian linearization [35], feedback linearization [35], Monaco and Normand-Cyrot's method [36] [37], etc. However, as mentioned previously not all biochemical pathway systems are subject to external cellar signals, therefore Jacobian and feedback linearization approaches which aim at design a complex input signal would not be discussed in this paper. In this section, we propose a time-varying linear approach based on Taylor expansion and Carleman linearization method for discretization of bilinear in the states pathway ODEs, and also briefly investigate Monaco and Normand-Cyrot's scheme.

1) Taylor-Carleman Method

We initially consider a generally nonlinear model of the form:

$$\dot{\mathbf{x}}(t) = \mathbf{f}(\mathbf{x}(t), \mathbf{\theta}) \tag{18}$$

Then using Taylor expansion around the current time instant $t=t_k$, the state value at the next sample point $t=t_k+T$ is given by:

$$\mathbf{x}(t_k + T) = \mathbf{x}(t_k) + \sum_{l=1}^{\infty} \frac{T^l}{l!} \frac{\partial^l \mathbf{x}(t)}{\partial t^l} \bigg|_{t=t_k}$$
(19)

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which can be further simplified as:

$$\mathbf{x}(k+1) = \mathbf{x}(k) + \sum_{l=1}^{\infty} \frac{T^{l}}{l!} \mathbf{x}^{[l]}(k)$$
(20)

As discussed in Section 2, for some bioinformatics systems, the nonlinear ODEs are simply bilinear in the states:

$$\dot{\mathbf{x}} = \mathbf{A}\mathbf{x} + \mathbf{D}\mathbf{x} \otimes \mathbf{x} \tag{21}$$

where \otimes denotes the Kronecker product and $m \times m^2$ matrix **D** is assumed to be symmetric in the sense that the coefficients corresponding to x_1x_2 and x_2x_1 are the same in value. For many biochemical pathway models, **D** is generally very sparse. So lets evaluate the first few derivative terms of (20) to deduce the overall structure of the exact discrete time model of (21): $\dot{\mathbf{x}} = \mathbf{A}\mathbf{x} + \mathbf{D}\mathbf{x} \otimes \mathbf{x}$

$$\begin{split} \ddot{\mathbf{x}} &= \mathbf{A}\dot{\mathbf{x}} + 2\mathbf{D}\dot{\mathbf{x}} \otimes \mathbf{x} \\ &= \mathbf{A}^{2}\mathbf{x} + (\mathbf{A}\mathbf{D} + 2\mathbf{D}(\mathbf{A}\otimes\mathbf{I}))(\mathbf{x}\otimes\mathbf{x}) + 2\mathbf{D}(\mathbf{D}\otimes\mathbf{I})(\mathbf{x}\otimes\mathbf{x}\otimes\mathbf{x}) \\ \ddot{\mathbf{x}} &= \mathbf{A}^{2}\dot{\mathbf{x}} + (\mathbf{A}\mathbf{D} + 2\mathbf{D}(\mathbf{A}\otimes\mathbf{I}))(\dot{\mathbf{x}}\otimes\mathbf{x}) + 6\mathbf{D}(\mathbf{D}\otimes\mathbf{I})(\dot{\mathbf{x}}\otimes\mathbf{x}\otimes\mathbf{x}) \\ &= \mathbf{A}^{3}\mathbf{x} + (\mathbf{A}^{2}\mathbf{D} + 2\mathbf{A}\mathbf{D}(\mathbf{A}\otimes\mathbf{I}) + 4\mathbf{D}(\mathbf{A}\otimes\mathbf{I})^{2})(\mathbf{x}\otimes\mathbf{x}) + \\ &\quad (2\mathbf{A}\mathbf{D}(\mathbf{D}\otimes\mathbf{I}) + 4\mathbf{D}(\mathbf{A}\otimes\mathbf{I})(\mathbf{D}\otimes\mathbf{I}) + \\ &\quad 6\mathbf{D}(\mathbf{D}\otimes\mathbf{I})(\mathbf{A}\otimes\mathbf{I}\otimes\mathbf{I}))(\mathbf{x}\otimes\mathbf{x}\otimes\mathbf{x}) + \\ &\quad 6\mathbf{D}(\mathbf{D}\otimes\mathbf{I})(\mathbf{D}\otimes\mathbf{I}\otimes\mathbf{I})(\mathbf{x}\otimes\mathbf{x}\otimes\mathbf{x}\otimes\mathbf{x}) + \\ &\quad (22) \end{split}$$

It can be seen that it is a polynomial in **x** of degree m+1. Hence the infinite sum in (19) is an infinite polynomial. We can notice that the coefficient of **x** in *n*th order derivative expansion is \mathbf{A}^n , and the coefficient of the second order terms $\mathbf{x} \otimes \mathbf{x}$ can be expressed recursively:

$$q^{1} = \mathbf{D}$$

$$q^{n} = \mathbf{A}^{n-1}\mathbf{D} + 2q^{n-1}(\mathbf{A} \otimes \mathbf{I})$$
(23)

The exact representation of (21) in discrete time should have the form:

$$\mathbf{x}(k+1) = \mathbf{p}(\mathbf{x}(k)) \tag{24}$$

where requires **p** to be a vector valued. Here, instead of treating the system as a global non-linear discrete time representation, it would be possible to treat it as a time-varying linear system, where the time varying components depend on **x**. It's obviously an infinite degree polynomial and some finite length approximation must be used instead. For instance, if we only consider the second order approximation of derivative terms in (22) and using *jth* order Taylor expansion, the discrete-time representation of (21) can be expressed as:

$$\mathbf{x}(k+1) = \mathbf{x}(k) + \sum_{l=1}^{j} \frac{T^{l}}{l!} \mathbf{x}^{[l]}(k)$$

= $\mathbf{x}(k) + \sum_{l=1}^{j} \frac{T^{l}}{l!} (\mathbf{A}^{l} \mathbf{x}(k) + q^{l} \mathbf{x}(k) \otimes \mathbf{x}(k))$ (25)

The advantage of this approach is it gives a finite polynomial discrete time representation for bilinear in states models. However, as shown in (22) the model expression would be complex when considering exact higher order derivative expansion, otherwise, the lower order approximation of derivatives have to be employed as in (25), and corresponding discretization accuracy would decrease accordingly.

2) Monaco and Normand-Cyrot's Method

In stead of approximating derivatives, a recent algebraic discretization method proposed by Monaco and Normand -Cyrot is based on Lie expansion of continuous ODEs. When considering nonlinear ODEs with the form (18), the corresponding discretization scheme can be expressed as:

$$\mathbf{x}(k+1) = \mathbf{x}(k) + \sum_{l=1}^{j} \frac{T^{l}}{l!} \mathcal{L}_{\mathbf{f}}^{l}(\mathbf{x}(k))$$
(26)

where the Lie derivative is given by

$$\mathcal{L}_{\mathbf{f}}(\mathbf{x}(k)) = \sum_{i=1}^{m} f_i \frac{\partial \mathbf{x}}{\partial x_i}$$
(27)

and the higher order derivatives can be calculated recursively

$$\mathcal{L}_{\mathbf{f}}'(\mathbf{x}(k)) = \mathcal{L}_{\mathbf{f}}(\mathcal{L}_{\mathbf{f}}'^{-1}(\mathbf{x}(k)))$$
(28)

Thus, (26) can be rewritten as:

$$\mathbf{x}(k+1) = \mathbf{x}(k) + T\mathbf{f} + \frac{1}{2}T^2 J(\mathbf{f}) * \mathbf{f} + \frac{1}{6}T^3 J(J(\mathbf{f}) * \mathbf{f}) * \mathbf{f} + \dots$$
(29)

where $J(\cdot)$ is the Jacobian matrix of the argument. This truncated Taylor-Lie expansion approach has been shown with accurate discrete-time approximation and superior robust performance especially when considering large sampling time step in discretization [37] [38]. However, this approach could also be computational expansive as a series of composite Jacobian matrices need to be calculated.

B. Multi-Step System Discretization

Runge-Kutta methods [25] which are widely used for solving ODEs' initial value problems can be a nature choice for discrete time system representation. Runge-Kutta methods propagate a solution over an interval by combining the information from several Euler-style steps (e.g. (30)), and then using the information obtained to match a Taylor series expansion to some higher order.

$$\mathbf{x}(k+1) = \mathbf{x}(k) + h\mathbf{f}(\mathbf{x}(k))$$
(30)

Here, h is the sampling interval, $t_{k+1} = t_k + h$.

The discrete time representation of continues-time pathway model (1) can be written as follows using Runge-Kutta method:

$$\mathbf{x}(k+1) = \mathbf{x}(k) + \mathbf{R}(\mathbf{x}(k))$$

$$\mathbf{y}(k+1) = \mathbf{g}(\mathbf{x}(k+1))$$
(31)

Here,
$$\mathbf{R}(\mathbf{x}(k))$$
 represents the Runge-Kutta formula.
According to desired modeling accuracy, different order
Runge-Kutta formula can be employed and corresponding

Runge-Kutta formula can be employed, and corresponding $\mathbf{R}(\mathbf{x}(k))$ would be different. For instance, the *second-order Runge-Kutta formula* with the expression:

$$d_{1} = h\mathbf{f}(\mathbf{x}(k))$$

$$d_{2} = h\mathbf{f}(\mathbf{x}(k) + d_{1}/2) \qquad (32)$$

$$\mathbf{R}(\mathbf{x}(k)) = d_{2}$$

where $\mathbf{f}(\cdot)$ is the right-hand side of model's ODEs in (1).

The most often used classical *fourth-order Runge-Kutta formula* is:

$$d_{1} = h\mathbf{f}(\mathbf{x}(k))$$

$$d_{2} = h\mathbf{f}(\mathbf{x}(k) + d_{1}/2)$$

$$d_{3} = h\mathbf{f}(\mathbf{x}(k) + d_{2}/2)$$

$$d_{4} = h\mathbf{f}(\mathbf{x}(k) + d_{3})$$

$$\mathbf{R}(\mathbf{x}(k)) = d_{1}/6 + d_{2}/3 + d_{3}/3 + d_{4}/6$$
(33)

In practice, fourth-order Runge-Kutta is generally superior to second -order due to four evaluations of the right-hand side per step h.

Compared with one-step discretization approaches discussed in previous subsection, the main advantage of using Runge-Kutta method for discretization is it utilizes multi-step information for approximation of one-step-ahead predictions. This strategy enhances the discretization accuracy and reduces the complexity of the mathematical expressions compared with using higher order derivative approximation in one-step discretization. Moreover, Runge-Kutta method could provide a general discrete-time ODE representation for either linear or nonlinear ODEs, whereas for one-step strategy using Taylor expansion it's difficult to formulate a close representation for nonlinear ODEs, instead some finite order approximation has to be used.

Similarly, now the parameter sensitivity equations (9) can also be discretized using Runge-Kutta method:

$$\mathbf{S}(k+1) = \mathbf{S}(k) + \mathbf{R}_{s}(\mathbf{S}(k))$$
(34)

Here within the Runge-Kutta formula $\mathbf{R}_{s}(\mathbf{S}(k))$, $\mathbf{f}_{s}(\cdot)$ represents the right-hand side of sensitivity equations (9), e.g.

$$\mathbf{f}_{S}(\mathbf{S}(k)) = J\mathbf{S}(t(k)) + P \tag{35}$$

Thus, parameter sensitivity coefficients (8) and Fisher information matrix (12) can now be represented and calculated iteratively using discrete-time formula (34).

In this section, two sorts with three kinds of discrete-time model representation methods are investigated for pathway ODEs in depth. The Runge-Kutta based method shows superiority in mathematical model expression especially for discretization of nonlinear ODEs. In next section, the simulation results of five discrete-time models based on these three approaches are discussed and compared numerically and graphically using Michaelis-Menten kinetic model.

IV. SIMULATION RESULTS

In this section, a simple pathway example is employed for illustration of different discussed discretization approaches. The accuracy and computational cost of different methods are compared as well. The example discussed here is the well known Michaelis-Menten enzyme kinetics. The kinetic reaction of this signal transduction pathway can be represented as:

$$S + E \underset{\theta_2}{\overset{\theta_1}{\longleftrightarrow}} ES \overset{\theta_3}{\to} E + P \tag{36}$$

Here, E is the concentration of an enzyme that combines with a substrate S to form an enzyme substrate complex ES. The complex ES holds two possible outcomes in the next step. It can be dissociated into E and S, or it can further proceed to form a product P. Here, θ_1 , θ_2 and θ_2 are the corresponding reaction

rate. The pathway kinetics described in (36) can usually be represented by the following set of ODEs:

$$\dot{x}_{1} = -\theta_{1}x_{1}x_{2} + \theta_{2}x_{3}
\dot{x}_{2} = -\theta_{1}x_{1}x_{2} + (\theta_{2} + \theta_{3})x_{3}
\dot{x}_{3} = \theta_{1}x_{1}x_{2} - (\theta_{2} + \theta_{3})x_{3}
\dot{x}_{4} = \theta_{3}x_{3}$$
(37)

Here four states x_1 , x_2 , x_3 and x_4 refer to *S*, *E*, *ES* and *P* in (36) respectively. As it is a set of linear in the parameter and bilinear in state ODEs, (37) can be written into matrix form:

$$\dot{\mathbf{x}} = \mathbf{A}\mathbf{x} + \mathbf{D}\mathbf{x} \otimes \mathbf{x} \tag{38}$$

Firstly, we consider one-step-ahead discretization methods discussed in Section 3. For the Taylor series and Carleman linearization based method, according to (20), the first and second order local Taylor series approximation can be expressed as:

$$\mathbf{x}(k+1) = \mathbf{x}(k) + T\dot{\mathbf{x}}(k)$$
(39)

$$\mathbf{x}(k+1) = \mathbf{x}(k) + T\dot{\mathbf{x}}(k) + T^2/2\,\ddot{\mathbf{x}}(k)$$
(40)

As shown in (22), here

$$\dot{\mathbf{x}}(k) = \mathbf{A}\mathbf{x}(k) + \mathbf{D}\mathbf{x}(k) \otimes \mathbf{x}(k)$$

$$\ddot{\mathbf{x}}(k) = \mathbf{A}\dot{\mathbf{x}}(k) + 2\mathbf{D}\dot{\mathbf{x}}(k) \otimes \mathbf{x}(k)$$
(41)

In this example,

We can notice that, for this simple kinetic example with only four state variables, matrix **D** ($m \times m^2$) is already in large scale.

For Monaco and Normand-Cyrot's method, here we employ a truncated third order Lie expansion for simulation. As shown in (29), we only need to notice **f** represents the right hand side expression of model ODE

$$\mathbf{f}(\mathbf{x}) = \begin{bmatrix} -\theta_1 x_1 x_2 + \theta_2 x_3 \\ -\theta_1 x_1 x_2 + (\theta_2 + \theta_3) x_3 \\ \theta_1 x_1 x_2 - (\theta_2 + \theta_3) x_3 \\ \theta_3 x_3 \end{bmatrix}$$
(43)

The general Lie series expansion (29) seems concise in expression, however, the composite Jacobian expression would be large in scale and computational costly.

Now we consider the multi-step discretization approach based on Runge-Kutta method. It is straightforward to formulate the second and fourth order Runge-Kutta discrete-time expression using (31)-(33). For this example, we just need to replace $\mathbf{f}(\cdot)$ with (43), and as all the states can be measured for this example, the measurement function \mathbf{g} is an identity matrix \mathbf{I}_4 , therefore:

$$\mathbf{y}(k+1) = \mathbf{x}(k+1) \tag{44}$$

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Comparing the discrete-time model expression using local Taylor-Carleman expansion, Monaco and Mormand-Cyrot's method, and Runge-Kutta method, the expression using Runge-Kutta method is more straightforward and compact, we only need to replace $\mathbf{f}(\mathbf{x})$ with the right-hand side expression of the specific biochemical reaction ODEs. On the contrary, the discrete representation using second-order Taylor expansion is already a bit complicated as the scale of \mathbf{D} matrix would become very large as the parameter dimension increases. As discussed in Section III. *A*, it would be even more difficult to formulate a model expression using third or higher order Taylor expansion.

The time series simulation using different discrete-time model expressions are displayed and compared in Fig. 1. Here, five discrete-time models, which are based on first and second order Taylor expansion, second and fourth order Runge-Kutta, and Monaco and Mormand-Cyrot's respectively, are employed for comparison. The initial values of states are: $x_1(0) = 12$, $x_2(0) = 12$, $x_3(0) = 0$, $x_4(0) = 0$. Parameter values are set to be: $\theta_1 = 0.18$, $\theta_2 = 0.02$, $\theta_1 = 0.23$. Simulation time period is from 0 to 10 with sampling interval 0.3. We solve the model's ODEs (37) using *ode45* function in Matlab with sampling interval 0.1, and suppose the result as an approximation of real system observation and to be a judgment of different discretization methods. The residual mean squared errors (RMSE) between different models' outputs and the observation are listed and compared in Table I.





Fig. 1 Time series simulation results of using five different discrete-time models

TABLE I

Time Series Simulation Residual MSE of Different Models					
	1st order	2 nd order	2 nd order	4 th order	Monaco &
RMSE	Taylor	Taylor	Runge-Kutta	Runge-Kutta	Normand-Cyrot
x1	0.4771	0.0205	0.0942	0.0306e-4	0.0020
x2	0.5054	0.0144	0.1019	0.0774e-4	0.0030
x3	0.5054	0.0144	0.1019	0.0774e-4	0.0030
x4	0.0503	0.0105	0.0020	0.1770e-4	0.0002
Total	1.5381	0.0598	0.3000	3.6249e-5	0.0083

Fig. 1 and Table I provide states' trajectory simulation results based on five different discrete-time models. It is clearly to see that the discrete-time model based on first-order Taylor series gives the worst approximation results with the largest RMSE, and the one based on fourth-order Runge-Kutta gives the closest simulation result to the real observation with the smallest RMSE. Besides, the discrete-time model using Monaco and Normand-Cyrot's method and second order Taylor series also give an acceptable approximation results with relatively small RMSEs to the real observation.

V. CONCLUSION

Quantitative discrete-time model representations are important as a link between continuous-time biochemical

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kinetic reactions and discrete-time experimentation. It will receive more and more attentions as computer based simulation and analyses are widely used in current biochemical pathway modeling study. Two important sorts of discretization methods are mainly investigated in this paper. One strategy is based on one-step-ahead Taylor or Lie series expansion. This kind of method could give an exact discrete-time representation for linear ODEs, however, for more typical bilinear or nonlinear ODEs pathway models, truncated finite order Taylor/Lie series approximation have to be used. The mathematical discrete-time expression using higher order Taylor/Lie expansion can be very complex and it would be computational costly as well. The alternative is the Runge-Kutta based approaches, which are multi-step discretization strategy. The mathematical model representation using this method is straightforward and compact, and the simulation approximation result using fourth-order Runge-Kutta is superior to others as well. Synthetically speaking, Runge-Kutta based discretization method can be a better choice for discrete-time model representation in pathway modeling study, and the corresponding discrete-time model structure will be a useful and promising tool in the future systems biology research. Further work can focus on dynamic analysis of discrete-time models and comparison with corresponding continuous model, here dynamic analysis should include model zero dynamics, equilibrium property, chaotic behavior when varying sampling step, etc. Additionally, discrete-time local and global parametric sensitivity analysis methods would also be a significant further focus for pathway modeling study.

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